## Name: \_

## **PHOTOSYNTHESIS LAB, Part 1 (procedure and data gathering)**

## **Procedure**

Almost everything you need to know for the set up of this lab you can get from watching <a href="https://www.youtube.com/watch?v=ZnY9\_wMZZWI">https://www.youtube.com/watch?v=ZnY9\_wMZZWI</a>

## What's below is for reference

- 1. Each group is going to set up three or four cups, each with 10 spinach disks. Use the hole puncher to punch these out. Avoid the leaf veins.
- 2. A solution of sodium bicarbonate (baking soda) with a tiny bit of detergent has been prepared for you in a beaker. Pour 100 mL of this solution into each of your clear plastic cups. Make sure there are no bubbles.
- 3. Remove the plunger from a large clean syringe. Place 10 leaf disks into the body of the syringe. Be sure the leaf disks are near the tip of the syringe as you reinsert the plunger so you don't damage the disks.
- 4. Insert the tip of the syringe into the beaker of baking soda solution and draw liquid into the syringe. The leaf disks should be floating at this time.
- 5. Hold the syringe tip <u>upward</u> and expel the air by depressing the plunger carefully. Stop before solution comes out the tip.
- 6. Seal the tip of the syringe using the thumb of your left hand and hold tightly. Pull back on the plunger creating a partial vacuum within the syringe. If you have a good seal it should be hard to pull on the plunger and you should see bubbles coming from the edge of the leaf disks. Hold for a count of ten.
- 7. Simultaneously, release your index finger and the plunger. Some of the leaf disks should start to sink. Tap the side of the tube or shake gently to break any bubbles on the edges of the disks.
- 8. <u>Repeat steps 6 and 7 until all the disks sink</u>. Don't overdo it, because that'll damage the leaf tissue.
- 9. Remove the plunger from the syringe and pour the solution containing the disks into one of the plastic cups. There should be 10 disks per cup. Make sure they sink to the bottom. Cover with a piece of paper so that they don't start to photosynthesize.

We'll have four experimental light treatments: 1) Red light, 2) green light, 3) blue light, and 4) white light. Your group will be assigned three of these four treatments.

10. Check your cups every minute for 10-20 minutes. At each time check, count how many leaf disks are floating at the top of solution in each cup.

PREDICTION: Based on what I know about PSN, I predict that:

DATA A	NALYSIS	How do we analyze these data?	Data table 1			Data tab	le 2	Data table 3				
SAMPLE DATA		Averaging wouldn't make sense.	Light treatment			Light treat	tment	Light treatment				
Minute # Floating		The best data point is to find the	(circle 1): red, green,			(circle 1): rec	l, green,	(circle 1): red, green,				
s	disks	point in time where 50% of the	blue, white			blue, wh	ite		blue, white			
1	0	disks are floating. In the data set to	Time (Min.)	# of Disks		Time (Min.)	# of Disks		Time (Min.)	# of Disks		
2	0	the left, that's at about 11.5		Floating			Floating			Floating		
Z	0	minutes. Why 11.5? Notice that at	0			0			0			
3	0	11 minutes there were 4 disks	1			1			1		1	
4	0	floating, and at 12 minutes there	2			2			2		1	
F	0	were /. That means that	3			3			3		1	
5	0	somewhere between 11 and 12	4			4			4		11	
6	0	minutes, half of the disks (5 of the	5		-	5			5			
7	1	This term is referred to as the $FT_{ro}$	6		-	6			6		1	
8	1	(which stands for <i>Effective Time</i> ,	7		-	7			7		1	
0	1	50%)	8		-	, 8			8			
9	1	The ET <sub>50</sub> has a problem though.	0		-	0			0		-	
10	1	The higher the rate of	9		-	9			9		-	
11	4	photosynthesis, the <i>lower</i> the ET <sub>50</sub>	10			10			10			
10	7	(because more disks will float in	11			11			11			
12	/	less time). Therefore, what we're	12			12			12			
13	8	going to figure out for each trial is	13			13			13		1	
14	10	the <i>inverse</i> of the ET <sub>50</sub> ., or 1/ ET <sub>50</sub> .	14			14			14		11	
L		You'll convert your fraction into a	15		1	15			15		11	
		decimal value. In the case above, 1/11.5 is 0.087	1/ ET <sub>50</sub> .			1/ ET <sub>50</sub> .			1/ ET <sub>50</sub> .			

	1/E <sub>50</sub> Class Means															
Data table 4	ſ				Red		Green				Blue				White	
Light treatment		Mean of 1/E <sub>50</sub>											-			
blue, white		Stan. Dev.														
Time (Min.) # of Disks Floating	-	Standar	d Err	or												
0		(for erro	or bai	s)												
1	Tiele			,												
2	Title:															т
3				_	$\square$				_							-
4																
5																
6																
7																1
											+		+			1
					+		_	+					+			1
					+						-					+
					+	+		+	_		-		_			+
			+ +		+	+		+			-		_			-
																_
$1/ET_{50}$ .													_			-
																_
After gathering your data,																
reassemble your lab kit,																
and post your data on the																
class After calculating																1
standard error, graph the													+			1
data above (four bars,					++						+		-			+
with error bars above					+				_		-		-			+
each one).			+		+				_		-		+			-
Formula for standard deviation Formula for standard error																
$\sum ( - )^2$						CE S										
$s = \sqrt{\frac{\sum (x - \bar{x})}{n - 1}}$						3	E =	1	n							

CONCLUSION: Explain what happened in this lab, and why results do or don't make sense. Include in your explanation appropriate references to the following concepts: the light reactions; absorption spectrum; action spectrum, oxygen production.